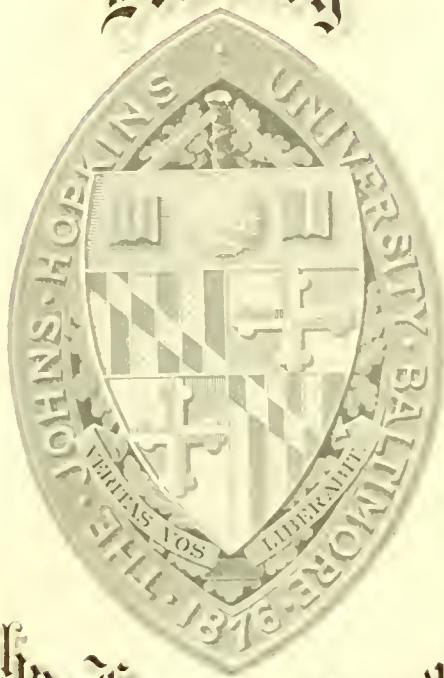


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ON THE DEVELOPMENT OF THE
LEAF AND SPOROCARP
IN MARSILIA QUADRIFOLIA, L.

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Dissertation presented to the Board of
University Studies of the Johns Hopkins University -
ersity for the degree of Doctor of Philosophy,

1911

Duncan S. Johnson.

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May 1, 1911.

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The Leaf and Sporocarp of Marsilia.

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Both genera of the Marsilieaceae have been the subject of frequent study by botanists since the early part of this century. But in spite of this the exact origin, sequence and development of the various organs of the mature sporophyte, and especially the morphological significance of the sporocarp, have never been made out satisfactorily in either genus. This is undoubtedly due in the main to the dense covering of trichomes, over all the young organs, which practically prevents the successful study of these parts in toto as transparent objects. But it is due partly also to the complexity of the apical bud with its confusion of numerous and crowded, root, branch, leaf and sporocarp rudiments, developing, except the last, in rapid succession from the segment of the apical cell of the stem. Still much has been accomplished even with this method and with hand sections, by Bischoff, Mettenius, Hanstein and Russow. The introduction of the paraffine method of sectioning has largely overcome the first difficulty, while solving the second remains more puzzling still. This method in the hands of Menier, Pfeiffer, Campbell and others, has added considerably to our knowledge of the minuter details of development in these forms.

The present work was undertaken at the suggestion of

Dr. J. P. Lotsy, of the Johns Hopkins University, for the purpose of determining the origin and early development of the sporocarp in Marsilia, in the hope that thus some light might be thrown on the question of its morphological nature. It was very soon decided, that the best way to reach this end would be to study in detail the origin and development of the leaf also, for comparison, and thus if possible to complete the detailed developmental history between the leaf mother cell and the sporangium.

The work has been carried on during the last two winters in the biological laboratory of the Johns Hopkins University, under the direction of Dr. J. E. Humphrey, to whom for his constant assistance and encouraging interest, I wish here to express my sincere thanks. I am indebted also to Dr. W. A. Setchell, then of Yale University, for aid in obtaining material.

The material used was obtained from New Haven, Conn. at Cromwell, Conn. It was fixed in either 95% alcohol, 1% chromic acid, or a sublimate-acetic mixture (5% of glacial acetic acid in a saturated solution of corrosive sublimate). All of these gave good results, but the sublimate mixture was the best on the whole, since the chromic acid specimens did not always stain well and at certain stages of development the alcohol caused shrinkage. The material was stained either in toto in Mayer's haemalum, or on the slide with this alone or in combination with Eisenmenger, with certain

2.

violet or alcoholic cocaine 1. Of these the latter with haemalum with Bismarck Brown gave the best results.

THE DEVELOPMENT OF THE LEAF.

Practically all that we know of the development of the leaf in *Marsilia*, is due to the work of J. Harstein, on the leaves of the young embryo, and it will be necessary to refer to this often as we proceed.

The leaves of *Marsilia* arise in two rows, one on each of the dorso-lateral surfaces of the stem. Each leaf arises from one of the cells derived from a dorso-lateral segment of the tetrahedral apical cell of the stem. The leaf mother cell is first recognizable when the segment in which it arises, is about the third or fourth in its series from the apical cell. It is soon distinguished by its larger size, usually by a larger nucleus and by its bulging beyond the general surface of the stem apex, (L. Figs. 1. 2.).

The first division of the leaf mother cell, is usually by a curved longitudinal anticline, appearing toward the dorsal edge of the cell and with its concavity facing laterally, (S. M. Fig. 2.). The second wall is also a longitudinal anticline near the lateral edge of the mother cell and with its concavity facing dorsally, (S. M. Fig. 1). These walls are also inclined to each other, (Fig. 2.) and there is thus left between them the typical two-sided anticlinal wall.

of the leaf, curved like the half of a circle convex below, and with its edges directed respectively toward the base and apex of the stem.

Hensteink ('05) has already shown that the leaf of *Marsilia* has a two-sided apical cell, and he figures it (Pl. XIII fig. 17) as having approximately the position given above, but he did not describe the origin of the apical cell except in the leaves of the very young embryo. This agrees in origin, shape and position with the apical cell of the leaf as found in most of the other Leptosporangiate Ferns that have been studied. They have been thus described by Hofmeister ('62) in *Aspidium*, by Strasburger ('73) and Campbell ('82) in *Azolla*, by Kny ('75) in *Ceratopteris*, by Klein ('87) in *Polypodium*, by Bower ('89) in *Trichomanes*, and by Campbell ('87) in *Onoclea*. In *Pteris* however, while the origin and shape of the apical cell are the same, its position, as shown by Hofmeister ('62) and Klein ('84) is exactly at right angles to that found in the other genera mentioned. That is the edges are directed transversely to the stem and segments are cut off toward and away from the stem apex alternately.

In *Pilularia*, the only other genus of the Marsiliace, Bower ('89) and Menier ('87) describe the apical cell of the leaf as two-sided, while Campbell ('83 pl. 12, '87 p. 11) states that it is tetrahedral, as differing from that of

Marsilia. The situation is the same as in Marsilia, the median leaves flat in other than the position of a green leaf entirely with Marsilia and thus with the other members of the group except Pteris.

The two sided apical cell thus formed in Marsilia, continues its growth and activity, cutting off segments alternately towards the dorsal and lateral sides of the stem (Fig. 3a), or, since the ventral side of the young leaf looks towards the apex of the stem (Fig. 3), the segments are cut off alternately toward the right and left of the leaf itself. This goes on until about fifteen or sixteen segments have been formed on each side, when the apical cell ceases to function as such and apical growth ends. The exact fate of the apical cell was not made out in the leaf, but it is probable that a periclinal wall is finally formed instead of the usual segment wall, as such a wall was seen several times in the apical cell of the sporocarp. Sadebeck ('73), Karr ('77) and Bower ('84) have actually observed such a wall in the apical cell of the leaf, but they were not able to make out just how many segments were cut off before it appeared.

In Marsilia the great regularity of the divisions in the segments of the leaf, as well as the fact that certain cells remain of the full length of the segment (Fig. 5), make it possible to determine quite exactly the number of segments cut off (Fig. 4). The only doubt is in regard to

the first segments, which may be the tip of the stem, so that it is not clear if that the second segment in Fig. 4, may not really be the second one. Aside from this the several leaves examined in which apical growth has been observed very closely as to the number of segments formed.

The young leaf at the end of segmentation of its apical cell, is about 1mm. long and .15mm. in diameter at the base, and nearly up to this time it has the form of a slightly tapering cone, capped by the bulging apical cell, which curves upward and ventrally (Fig. 4) over the stem apex, by more rapid growth on the lower and dorsal sides. The cross section of the leaf at this state is almost exactly circular. The leaf not being, until the formation of the pinnae, at all flattened or ^{or} crumpled, as described by Campbell ('85) and figured by Hanstein ('66) in *M. Drunnondii*.

Just before the last few segments are formed, the tenth and eleventh, or eleventh and twelfth, segments from the base on each side, begin to swell out laterally and ventrally to form the first pair of pinnae of the lamina (n^o Fig. 31). Soon after the last segment is cut off from the apical cell, the segments beyond the first pair of pinnae develop the second and terminal pair of leaflets (n^o Fig. 32). If the leaf bears one or more sporocarps, the first of these arises on the lower and anterior side of the petiole from the second segment on its side from the base (Fig. 22). The second

sporocarp is present we also find from a segment of the petiole near the second, which usually arises on the first sporophyll near its base, and on the side turned toward the petiole.

The segments of the apical cell of the leaf, or primary marginal cells of Hanstein ('65) and Sadebeck ('71), are quite regular in shape, being curved slices, slightly thicker at the middle of the outer border (Fig. 6) and with a slightly curved inner border where the segments overlap (Fig. 8). The first division wall to appear in these marginal cells is a longitudinal and radial enticline (Fig. 9), running from about the middle of the inner border to the dorsal part of the outer border, so as to cut off about a third of the whole semicircular segment toward the dorsal side (I, Figs. 7 - 10). We may call the third thus cut off a section, as the part still left of the primary marginal cell is the secondary marginal cell (m.c.², Fig. 9). Wall I is apparently the "tangential wall" of Sadebeck ('74), and section I is the "schichtzelle" of Hanstein ('65). But this terminology does not seem appropriate here, when the real position of this and the later section walls is taken into account, since it refers to the position of the walls at the surface of the leaf only, when the third of real importance is their position in the interior, as we shall see.

The second wall formed in the segment is located in-

itudinal anticline (II Figs. 6, 10), but instead of this radial it is nearly parallel to the inner or median order of the segment, running from wall I to the ventral outer border of the secondary marginal cell, thus cutting off section II and leaving a tertiary marginal cell (t.m.c.³, Figs. 10, 11). In each of these sections and in the tertiary marginal cell walls are formed in various planes nearly simultaneously as the leaf grows. In section I there is formed first a pericline (pl. w., Fig. 11), at about one third of the distance outward from the center of the leaf, cutting off thus at the inner end a part of the plerome contributed by this section to the axial bundle of the leaf. Then a longitudinal and radial anticline, which we may call the halving anticline, cuts the outer cell into two (h.a., Figs. 7, 11). Meantime the plerome of section II is cut off by a pericline (Fig. 11), while the tertiary marginal cell divides by a transverse anticline (t.a.), Figs. 6, 7) into an acrosopic and a basiscopic marginal cell. This anticline is the "radial wall" of Sadebeck.

The further divisions can best be described by taking up first the fate of the two tertiary marginal cells. In each of these there is formed first an anticline (III, Fig. 11) nearly parallel to wall I and toward the dorsal side of the marginal cell; then a similar wall is formed toward the ventral side parallel to wall II (IV Figs. 8, 11'). There

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are thus left two marginal cells of the fifth grade in each segment. The next wall formed in each of these may have either one of two positions. In the more frequent type of division, another longitudinal anticline appears in each (V, Fig. 13) parallel to wall III, forming thus the fifth and last section and leaving an ultimate marginal cell of the sixth grade. Then a pericline appears, running from wall IV to wall V, and the function of the marginal cell as such is thus ended. In the other and less frequent type of division, the marginal cells of the fifth grade are divided immediately by periclines running from wall III to wall IV, the marginal cell of the fifth grade playing the same role here as that of the sixth grade in the first type; and their further development is also practically the same in both cases. The fate of all the segments of the leaf is in other respects the same, with the exception of those giving rise to the lamina and to the sporocarp, in which the pericline is never formed in the marginal cell.

THE RETIOLE.

Taking the first type of leaf for the study of the further development of the segments of the retiole, we find that during the divisions of the tertiary marginal cells, sections I and II, and the other sections as fast as formed,

so on dividing successively. Section I cuts off a dermatozen layer by a pericline near the outer end of each half, then a second plerome portion is cut off from the inner end of each (d. and pl., Fig. 13) and section II, followed by each of the others in turn, cuts off plerome and dermatozen (Figs. 13 - 15), while the ultimate marginal cell is last of all to form, first the dermatoren wall as described above, and then still later the plerome wall (pl. w., Fig. 16). The dermatoren soon splits by a pericline into two layers around the whole circumference of the petiole, and these break up by anticlines, both longitudinal and transverse, to form the epidermis and hypodermis, each of which remains of one cell in thickness even in the mature leaf (en. y. Figs. 14 - 19). On the line of the median wall, of each of the section walls, and of the halving anticline of section I, small intercellular spaces soon appear between the cells of the periblem and those of the hypodermis (e.c., Figs. 13 - 19). These are the beginnings of the longitudinal air canals of the petiole, of which, as can be seen from their origin, fourteen are formed altogether in leaves where the segments divide according to this type.

The single periblem cell formed in each half of section I, in each of the other sections and in the ultimate marginal cell (pl. Figs. 12 - 15), soon divides by a pericline into two cells (Fig. 15). Of these the inner cell di-

sides of anticlines and periclinal to form the mesophyll cells of the future leaf (m., Figs. 1^c - 19), while the outer cell gives rise to both the longitudinal and the transverse partitions between the air canals (r.c., Figs. 1^c, 1^e). These latter cells swell out in the middle (Figs. 16 - 18) and grow out at the ends into papilla like tips, which are in contact with those of their fellows of the adjacent sections (Figs. 17, 21), leaving an intercellular space between these and the mesophyll cells and also one above and below each papilla, connecting this with the primary air canal formed between the partition cells and the hypodermis (Figs. 16, 17, 21). These papillae are soon cut off by longitudinal anticlines (Figs. 17, 21 - 25), and thus is formed a pair of small nearly isodiametric cells, in each canal opposite each of the primary partition cells, of which there are usually eight in the length of a segment. From these eight pairs of cells, one cell in each pair being from each of the opposite primary partition cells bordering on an air canal, are developed the eight transverse partitions of the air canal in each segment. These remain one cell thick even till maturity, but later in their development many intercellular openings or pores are formed, allowing the passage of air through them (e.g., Figs. 1^c, 20, 21). Except for these pores, the partitions of the mature leaf stretch completely across the entire

parallel to the odd one in the cross partition in which position it is in the part.

That part of each primary partition cell left after the separation of the single cross partition cell at each end, does not divide further by longitudinal anticlines, till very much later, but immediately gives rise to the longitudinal partition between the adjacent air canals. As these cells grow in a radial direction they are divided by periclines until, in the mature leaf, the partitions may be fifteen cells or more in radial breadth and four across the diameter of the petiole, but they are only one cell thick (L.p., Fig. 18, 20). These cells grow with the longitudinal growth of the leaf (Figs. 11 - 21). As each cell elongates it is seen that the primary cross partition cell at one end (c.p.c., Fig. 20) is nearer the acrosopic wall of the longitudinal partition cell, and to the other end is nearer the basiscopic wall. Then when the transverse anticline appears it is somewhat oblique, and thus forms two wedge shaped cells, each with a cross partition cell at the broad end and none at the narrow one (Fig. 22). The cross partitions in adjacent air canals opposite is no opposite, but alternate. These wedge shaped cells continue to grow and divide frequently by transverse anticlines, till in the mature petiole the cross partitions are far apart. Here in the longitudinal partitions also

find at maturity, with small intercellular canals so-called "nerves" of Münker, which allow communication between adjacent canals (L.P.P., Figs. 19, 20, 22).

In the type of petiole with five sections in each segment, there are, as we have seen, fourteen primary air canals. When the segments of both sides of the petiole form only four section walls, there are but twelve canals, while when, as occasionally happens, one side forms four and the other five sections, we find thirteen canals. All three types are found in the older petioles, but it is interesting to note that the only figure of a cross section of a petiole that I have seen, that of Bischoff ('23), is a petiole of the frequent type with thirteen canals.

At a comparatively late stage of the development of the leaf, when the longitudinal partition is already many cells broad radially, the outer cells, next to the hypodermis, or the inner ones next to the mesophyll cells, may split by a longitudinal anticline. The daughter cells then separate laterally (a.c., Fig. I), and divide by periclinics, forming thus a new canal bordered by these cells and the hypodermis or meso and the mesophyll cells. This splitting of the primary partition may apparently proceed through its whole radial width, and give rise to a secondary canal between the two layers, largely to be distinguished from the primary canals in the mature leaf. The number of the sec-

Outer canals thus formed is always small, the number of the primary ones, but there is apparently no regularity either in number or position.

When the epidermal layer of a section has divided by one longitudinal anticline and three transverse ones (Fig. 27), forming thus eight cells altogether, four in the length of a segment and two in the width of a section, there is cut from each of these cells, by a circular anticline at the acroscopic end, a small cell which gives rise to one of the numerous trichomes that clothe the young leaf. The rest of the original epidermal cell then divides to four cells with the trichome at the acroscopic end of one of the upper pair (Fig. 28), then further divisions follow and at maturity the epidermal cells become much elongated (Fig. 29), as has been shown by Russow ('72) and also by Meunier for Pilularia, while some of them may later form more secondary trichomes. Each trichome cell grows out beyond the surface of the epidermis (Fig. 26) and swells at its outer end to a knob, which soon elongates, in the direction of the length of the petiole. On the basiscopic side it projects but slightly, while it grows out toward the tip of the leaf into the long multicellular hair, which is supported by the basal or stalk cell that remains wedged in between the other epidermal cells (Fig. 30). Before the leaf reaches maturity these hairs separate

from four pairs of collenchymatous longitudinal fibers. The later development and mature structure of these sections have been studied by Russow, and have been worked out in great detail by Meunier for the similar ones of *Pilularia*.

Stomata also occur on the petiole, but apparently not till quite a late stage, and their development was not studied.

By the time that the longitudinal partitions are two or four cells in radial width, a longitudinal row of the large mesophyll cells opposite each partition, usually in the next to the inner layer of these, becomes specialized to form one of the so-called tannin sacs (I.s., Figs. 11, 12). The contents of these are distinguished by a brownish color in the unstained alcoholic material, but usually stain very darkly with haemalum. In the mature leaf their cells are about the same size as the surrounding mesophyll cells. The latter are about twice as long as broad and fit together quite squarely at the ends, while laterally they are rounded off from each other, leaving between them many small intercellular spaces (Fig. 20) which connect with the large air canals.

While these structures have been developing from the dermatogen and periblem layers of the petiole, the plerome portions of the first four sections have given rise to the axial vascular bundle. The first plerome cell cut off from

section I, dividing the cells of the segment from a primary portion of the section, divided by many periclinal and longitudinal anticlines (Figs. 1, 12 - 15). Later transverse anticlines appear, the first one being in about the tenth segment from the apical cell of a leaf of the size shown in Fig. 1. In section II a longitudinal anticline is formed in the plerome cell parallel to the median wall; then a similar wall perpendicular to this divides the cell one into quarters (Figs. 16 - 17). Of these, the cell in the angle between sections I and III (Fig. F, 18 - 19) never divides further, but becomes directly the large trachea of its side of the axial bundle. Though no division walls are formed in this its nucleus divides actively, so that in a trachea of half a millimeter in length (Fig. F), there may be twenty-five or thirty nuclei present. Later these and the other protoplasmic contents of the trachea disappear, while the end walls, which are portions of the original segment walls, assume gradually the oblique position characteristic of these in the mature trachea (Fig. G). Their inclination is always downwards and dorsal as shown in the figure. This is the only cell of the segment that remains of the full length of a segment for any considerable time during the development.

The other three quarters of this section, and all the

plerome of sections III and IV divide up like the plerome of section I, by frequent periclinal and longitudinal anticlines, and by fewer transverse anticlines, to form with this the remaining tissues of the axial bundle. The small cells of the outer layer of the bundle gradually become more regular in shape and uniform in size, forming thus the characteristic bundle sheath (b.s., Figs. 18, 19).

The plerome of the ultimate marginal cell never takes any part in the formation of the axial bundle, but in some cases that of section V apparently contributes to the formation of the sheath and of a few cells within this (Fig. 18).

The Lamina.

By the time that the apical cell of the leaf has ceased to cut off segments, the tenth and eleventh (or eleventh and twelfth) segments from the base on each side, have begun to grow out laterally and ventrally to form the first pair of pinnae (p!, Fig. 31). These pinnae do not correspond in extent or direction to the segment, as it is known to do in *Asplenium serpentini* by Sadebeck ('72) and in *Onoclea struthiopteris* by Campbell ('87). In this respect they agree rather with the pinnae of *Ceratopteris thalictroides*, as described by Kny ('70). The lower border

of a pinna (Fig. 31) may correspond exactly with a segment wall while the upper border falls short of the second segment wall above, or perhaps the reverse may be true though it was never seen satisfactorily. It is certain, however, that the lower pinna on each side is formed from the whole length of one leaf segment and the larger part of another, and never of a single segment or of more than two. After cutting off its last segment the apical cell of the leaf, as stated above, probably breaks up to form marginal cells like those formed in the segments. By the time that this has happened, the segments beyond the first pair of pinnae, except all or part of the one next to the pinna on each side, have begun to swell out (Fig. 32) to form the terminal pair of pinnae. Later on the marginal cells arising from the apical cell probably take part also in the development of these pinnae.

The beginning of the development of a pinna is best seen in a transverse section of the leaf through the pinna, and is practically the same for both pairs. While in the segments of the petiole there are only five section walls formed, and the marginal cell of the sixth grade is the ultimate one, in the segments giving rise to the lamina no perclinal (dermatogen and plerome) walls are ever formed in the marginal cells. These on the contrary continue to form walls parallel to the longitudinal section walls

ill late in the development of the lamina (Figs. 34, 35).

The next wall formed after wall V is toward the ventral side of the marginal cell (VI, Figs. 34, 35), continuing thus the regular alternation of the section walls. Then wall VII is formed toward the dorsal side of the cell, wall VIII toward the ventral side, and so on till a large number of sections, if we may still call them such, have been formed (Fig. 35), the marginal cell being still distinguishable, and probably active, when the pinnae are more than two millimeters broad. These additional sections divide like the earlier ones by periclines to form the plerome, periblem and dermatogen layers and later by anticlines (Fig. 35), forming thus the various tissues of the lamina. Meanwhile the marginal cells divide also by anticlines perpendicular to the edge of the pinna (Fig. 36). They thus constantly increase in number and form a rounded growing edge, as Hanstein ('65) has shown, the pinna finally becoming wedge shaped with a broad rounded outer end, in the mature leaf. The pinnae are directed more ventrally than laterally from the petiole even at the beginning, and those of the upper or younger pair soon come to have their ventral surfaces nearly in contact, while the lower and older pair are folded obliquely in the leaf bud so as to enclose the younger ones between them (Fig.).

A branch of the axial bundle of the pinna is given off to each pinna, and each of these, branches in the lamina to form the anastomosing veins characteristic of the leaves of Marsilia. The detailed development of the bundles of the pinna was not studied, and it is not certain whether they originate as Sadebeck ('74) has shown them in *Asplenium*. The epidermal cells of the leaf give rise to stomata on the upper side, or on both upper and under sides, and to deciduous trichomes like those of the petiole.

In the leaf of *Pilularia*, which was examined for comparison with *Marsilia*, the segments of the two sided apical cell form but three sections, and leave thus an ultimate marginal cell of the fourth grad. The sections correspond in relative position to the first three in *Marsilia*, but they and the ultimate marginal cell are broader than in the latter, since the whole semicircular segment is here made up of four divisions, instead of containing six divisions as in *Marsilia*. Here as in *Marsilia* the outer part of section I is split by a pericline in two cells, each of which plays the same role as one of the other whole sections, in the development of the parichnos and dermatogen structures. The marginal cell, as well as all of the sections, here takes part in forming the axial bundle, and as will be seen from the number of sections, there

merous and primary tracheæ formed in section I, as in figure 1, Pischoff ('88, VIII, 1) and Campbell ('90, XV, 10). The partitions like the sclerites from which they arise are thicker, and seem to be more irregular in both size and position, than in Marsilia. The trachea formed in section II of each segment is also less prominent from its size than in the petiole of Marsilia.

The Sporocarp.

The bean-shaped sporocarps of *Marsilia quadrifolia* are most often borne in pairs, the stalks of the two apparently uniting below to form a common stalk, which joins the petiole of the fertile leaf on its inner side near the base. Occasionally only one sporocarp is found on a leaf, or there may be two with their stalks inserted separately on the petiole, and these may be several millimeters apart at maturity. Again, more rarely still, there may be three or four sporocarps, all with a common stalk, or one may arise separately from the petiole while the others are joined to it by a common stalk. In the former case, when the sporocarps are not yet full grown, it is usually found that the smaller or younger one of a pair, or of three, is borne on the side of the stalk toward the

Sporocarps on older leaves.

It is usually found that when no sporocarps occur on a branch or the stem, they are present on practically all the leaves of this branch. This fact was of very great assistance in the study of sections of the buds, or slightly developed branches, since the absence of sporocarp rudiments on the older leaves of a bud enabled one to avoid a fruitless search for the less easily recognizable earliest rudiments, on the younger leaves of the same bud.

It is an interesting fact that of the several localities from which material was obtained, the plants from those habitats where the water level lowered considerably to the end of the growing season, leaving the plants to grow on the wet mud and in the air, matured many more sporocarps than those which were submerged throughout the entire growing season. This appears not to be due to the fact that fewer sporocarps arise on the leaves of the submerged plants, but rather to the fact that the rudiments which are formed do not complete their development. At the end of July many young sporocarps are found on plants from either habitat, while at the end of September plants that have been left out of the water, by the fall of the water level, have many mature sporocarps on them and those that are still submerged have very few. Close examination of the leaves of the plants

in. In this, however, both the sterile and fertile
or non-ligulate leaf, and in the more or less developed
size which have evidently been arrested in their development,
and appear shrunken or withered. The leaves on which
these are borne have gone on in their development, and
we may thus find on full grown leaves sporocarps of the
same size as those found on other leaves of which the
pinnae have not yet unfolded. There is thus, on the
whole, no great regularity in the retardation in development
of the fertile or sporocarp bearing leaves.

Of the origin of the sporocarp of *Marsilea*, Bischoff ('28) says, that it arises as a slight prominence or papilla on the anterior side of the base of the petiole. Mettenius ('41) on the other hand, states that it arises endogenously and later breaks through the epidermis of the petiole, to form a projecting solid mass of tissue in the interior of which the various internal structures of the capsule are formed. The youngest sporocarps found by Russo ('72), had a two sided apical cell, but were already differentiated into sterile stalk and a fertilis-
tip or capsule, being probably in about the same stage as that shown in Fig. 42. He traced the development of the soral canals, stating that they arise by the splitting apart of certain cells in the interior of the capsule, and the development of pits on the ventral surface of the

in which the spores will open, leaving the soral canals but opening in order to close, to be closed again later by the growing together of the cells of the ventral surface. On the outer walls of these cavities arise the "soral cells", in each of which walls appear later so as to cut out a tetrahedral apical cell. This cuts off a number of segments, which according to Russow give rise to the placenta with its vascular bundle, and to the microsporangia, then a pericline appears at the inner end of the apical cell forming thus the archesporium of the single macrosporangium arising from it. Goebel ('62) states that the soral canals of Marsilia are external in origin and that the sporangia arise from superficial cells. Busgen ('90) described the earliest stage of the sporocarp as in one case a "gap" (Lücke) in the epidermal tissue of the young leaf near its base, and in another place as a swelling or prominence in the same region. The sporocarp grows for a time by a two sided apical cell, and he thinks it probable that all of the soral cells are derived from an epidermal cell of the ventral side of the capsule. The placenta macrosporangia and microsporangia he states are all formed as Russow has shown from these soral cells.

In the similar sporocarp of Pilularia, Holmeister ('62) and Juranyi ('79) describe the soral cavities as arising internally, while Goebel ('62), Meissner ('77) and

Campanile (1885) says nothing on this point, and in his
Goetzel and Rössler had found no such case in Marsilia.

According to my own observations on *Marsilia quadrifolia*, the sporocarp first makes its appearance on a young leaf on which it is borne, consists of about six or seven segments on each side; thus long before the lamina, or even the segments that are to develop this, have been formed. It arises by the increase in size and bulging out of apparently either the acrosticopic or the basiscopic ultimate marginal cell, of what is probably the second segment on its side from the base of the leaf (Fig. 37). Because of the beginning of curvature of the leaf, the proximity of the sporocarp rudiment to the axillary bud of the same leaf and the resulting difficulty in orienting so as to get exactly longitudinal sections of the leaf through the sporocarp rudiment, it was impossible to decide certainly in which segment the latter arises. It is probably the second rather than either the first or third segment, and cannot be a younger one than the latter in any of the cases seen, and though no indication of this was seen, it is possible that the exact point of origin may be shown, on further observation to vary. The sporocarp always arises from a marginal cell of the inner and ventral side of the young petiole (Fig. 3), the tip of which at this time is beginning to take a position nearly

parallel to the stem (Fig. 1). Soon however in the sporocarp mother cell, a curved anticline runs parallel to either its acroscopic or its basiscopic wall, then a similar transverse anticline on the opposite side of the mother cell (Figs. 27, 30), and there is thus formed a two sided apical cell right of the leaf, with its edges directed across the leaf. The sporocarp, as it arises thus from the whole of a marginal cell, which has not yet given rise to the three meristem layers that it is capable of forming, is not, strictly speaking, epidermal in origin. In its origin, by a two sided apical cell formed in a marginal cell of the leaf, the sporocarp resembles closely the solitary sporangium of *Lycopodium*, which as was shown by Prantl (Sadebeck '82, Fig. 64) arises from a marginal cell of the fertile pinnule.

The apical cell of the sporocarp thus formed goes on cutting off segments alternately toward and away from the leaf apex, which are to form the right and left sides of the sporocarp, till more than twenty have been formed on each side. It thus gives rise to a slightly tapering conical structure, much like the young leaf, which bends laterally to grow up beside the leaf (Fig. 1) with its ventral side facing in the same direction, but soon begins to bend ventrally upon itself (Fig. 47). Finally about the time that apical growth ceases, the upper part of cap-

sterile line with its ventral side nearly in contact with that of the lower or sterile portion of the sporocarp (Fig. 43). There is never any curling ^{of} in the extreme tip of the sporocarp suggesting the circinate coiling of the leaf, and the bending mentioned is later partially straightened out, as Russow ('72) has shown by the more rapid growth of the ventral side of the capsule. The activity of the apical cell as such, is ended by the appearance in it of a wall, which in a horizontal section is apparently a periclinal, but its exact direction was not made out satisfactorily in sagittal sections. At the time when this wall appears the bending mentioned above has taken place, the capsule is a millimeter long, and the sori at the base of this are developed as far as that shown in Fig. 10.

The first few pairs of segments of the sporocarp, probably four or five, form the sterile stalk, and then develop the fertile tip or capsule. Of the segments of the capsule the first three pairs do not develop sori and only the youngest one forms a lateral bundle. Each segment of the next eight or nine pairs gives rise to a single sorus and to a lateral branch of the dorsal bundle of the capsule. Beyond the last of the soral segments are six or seven pairs, of which only the three or four oldest form lateral bundles and none to form sori. The capsule

soon becomes flattened (Fig. 11), and during its early development is closely appressed to the trichomes (Fig. 12) and thickly covered by the trichomes arising from the capsule itself and from the stalk of the sporocarp. These trichomes have been omitted in all drawings because if they had been drawn they would have hidden the more important details in many of the figures.

The shape and size of the segments cut off from the apical cell here, are very nearly like those of the leaf, and their early divisions are exactly the same. Section walls I and II (Fig. 14) are formed in the same position as in the segments of the leaf, then the transverse anticline appears in the marginal cell (*t.a.*¹, Fig. 15) forming two of these of the third grade, and section wall III is formed in each of these in the usual position (III, Fig. 14). Thus up to this stage the series of figures given for the leaf (Figs. 7 - 11), is equally suitable for the sporocarp. After the formation of wall III, however, the regular alternation, that continues in the leaf till the last section wall is formed, is here broken by the appearance of wall IV on the dorsal side of the marginal cell and parallel to III, instead of on the ventral side as in the leaf (IV, Fig. 15). Wall V in the sporocarp is then formed toward the ventral side of the marginal cell, in the position of II

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IV of the leaf segments (V, Fig. 4), and so on up.
This is the last section of the leaf (VI, Fig. 10) is formed near
the dorsal side of the marginal cell. The ultimate marginal
cell is thus of the seventh grade here, and not of
the sixth grade as in the leaf.

When a second sporocarp is formed on a leaf, it usually
arises from a marginal cell of the second, or third seg-
ment of the first sporocarp on the side of the latter
that is turned toward the petiole which bears it (Fig. 10), and a third sporocarp may probably arise from the
second in the same way. The exact origin of the apical
cell here in the marginal cell was not seen but it is
reasonable to expect it to be like that of the first spo-
rocarp in the marginal cell of the leaf, and the earliest
^{stages} also indicate that this is true. This origin of the
younger sporocarps from the older ones, explains the occur-
rence of the mature sporocarps in pairs or trios on a com-
mon stalk, that was mentioned above. But the portion of
the stalk of a pair that is common to both is not, as sug-
gested by A. Braun ('70), formed by the fusion of two o-
riginally separate stalks, but it is the lower portion of
the simple stalk of the older sporocarp, and so of the com-
mon stalk of the second and third sporocarps of a pair.
Where two sporocarps are inserted on the petiole a com-

arate stalk, in which the two walls arise from
marginal cells of the stalk, and this is a comparatively
rare occurrence and was not actually seen in a well
staged colony of the alga.

We may return now to the further development of the
segments of the sporocarp. The positions of section walls
as given above is in general that found in all the seg-
ments, but there are frequent exceptions to this in various
segments that are worthy of note. Thus section V is usu-
ally broader in the acroscopic marginal cell of the soral
segments than in the basiscopic cell (V, Figs. 49, 117).
Sometimes section IV is also slightly broader and thus
since section VI does not vary greatly in size, the basi-
scopic ultimate marginal cells, which are evidently the
"sorus mother cells" of Büsgen, are the largest ones of the
whole sporocarp. It quite frequently happens also that
wall IV when formed, in the acroscopic marginal cells
of the soral segments and in either marginal cell of the
other segments, may bend down at the inner end and intersect
wall III instead of running through to meet wall
II as usual (IV, Fig. 4). This was frequently seen
in the younger stages as figured and in later stages in
the segments of the stalk but was never seen in the basi-
scopic marginal cell of the soral segments. Again, the final
cell of the sporocarp except one basiscopic one

more or less than the size of a pin-head, and will be
noticed in the small V-shaped depression which occurs
in the sixth or the seventh row. This is apparently
analogous to the behavior of some marginal cells of the
fifth grade in the leaf.

The Stalk.

If we follow out the development of each of the sections of the different regions of the sporocarp, we may best begin with the four or five pairs of segments of the stalk. We shall find that these together produce at maturity a structure much like the petiole of the leaf, or about half a millimeter in diameter and from ten to twenty millimeters in length. Each of the sections of these segments cuts off plerome and dermalogen cells, and section I cuts off a second portion of plerome as in the leaf, but only the plerome of sections I and III, and still only part of section III (Fig. 0), take part in forming the axial bundle of the stalk. The plerome of the other sections, together with the portion of all the sections, develops into a thick wall of somatic cells, if we may call them such, which give the mature stalk its rigidity and strength, a condition which pro-

and the arrangement of the mesophyll follows the lower part of the diagram, and the base of the older capsule (i.e., Fig. 1). The dermatogen lies close to the pericicle of the hypodermis, and to the epidermis with its trichomes and scales, while between the hypodermis and the pericile are found the small and irregular air canals. The number of them more than twenty of them, and six on each side, is seen between the pericile and hypodermis of sections I and III. It is worthy of note that the dermatogen of sections II and V soon splits by longitudinal anticlines (Fig. 50) as was seen in the leaf (Figs. 15, 16), and differs from the dermatogen of the other sections in the smaller segments of the capsule, as we shall see later. None of the mesophyll cells either here or elsewhere in the gynoecarp become specialized as tannin reservoirs as we have seen them to in the leaf.

The Capsule.

Turning now to the development of the second series of transverse sections of the capsule, we find that in section IV the plerome in dermatogen as in the stem, in the pericile between (pl., p. 2, d., Figs. 11-13, 51), but only in the plerome portions of section I it appears in form of small rounded cells scattered in the

size (..., p. 1). Several of the cells of the different portions of section I show the following arrangement of the segments: no one cell is found to contain more than two segments. The longitudinal vascular bundle of the sporocarp is thus very much more localized than that of the leaf, which has been seen to arise from the apices of four of the sections. The dermogen of all the sections in all the segments of the capsule splits up in the leaf into epidermal and hypodermal layers. Of these the former remains, as in the leaf, of one cell in thickness and gives rise to trichomes and stomata. The latter has been shown by Russo and in Pilularia by Meunier, divides (hy., Figs. 5a, 5b) to form the two layers of thickened cells of the wall of the mature capsule, and differs thus from the one layered hypodermis of the petiole (Fig. 10). In general the periblem of all the sections, in all the segments, gives rise to the few layers of loosely packed cells between the vascular bundles and the hypodermis. Between these cells and the hypodermis are formed the small and irregular air canals, which in the capsule are confined largely to the dorsal side and thus to sections I and III (a.c., Fig. 51).

It will be best now to examine in detail first the further development of the eight or nine somatic segments of the young basal portion of the plant, or prothallus.

We have seen above that section I in the stalk of the petiole, as well as in the other segments of the sorus, develops much as in the specimen of *Hypnea* of the petiole except for the peculiar specialization of the hypodermis. All the other sections on the contrary have a peculiar history differing from those of either the stalk or the leaf. Sections III, IV and V, dorsal to the marginal cell, widen rapidly at their outer ends, and, as sections II and VI do not widen in this way, the ultimate marginal cell is pushed into a ventral position (Figs. 45, 46, 51). Of course the interpolation of the extra section IV, dorsal to the marginal cell, would give the latter a more ventral position than it has in the petiole (Figs. 17, 18), and this is evident. The use of its formation out of the normal sequence, but this result is greatly enhanced by its very vigorous expansion at the outer end. Finally section VI grows out at the ventral end, beyond the cells arising from the basiscopic marginal cell and spreads over into contact with the cells of section V at the ventral surface (Figs. 52, 53). The plerome of the basiscopic or soral half of section III breaks up by one anticline and then by periclines (Figs. 51, 55) to form the dorsal part of the lateral band. The more dorsal part of section IV does likewise, while

in the more ventral part, as is also later, the fibers of the lateral bundle arise ^{in the} from the oscopic or inner halves of the segments (l.b.f. Fig. 4). In section VI the plerome of the basicoptic half gives rise to the lateral bundle of the sorus and to the branch connecting this with the lateral bundle (pa.b., pa.b., Figs. 48 - 50).

Of the sections on the ventral side of the marginal cell, the plerome of section II divides to form rather large isodiametric cells (pl. Figs. 51, 54) which ultimately form the gelatinizing tissue of the dorsal part of the capsule described by Hanstein ('82) and Russow ('82). The plerome of section V divides by a longitudinal and radial anticline (l.a., Figs. 47, 51) and grows around under the inner end of the cells of the sorus, and the inner end of section VI (Figs. 51 - 54). The ultimate fate of the plerome of this section is probably to help form the dorsal part of the gelatinous ring along with that of section II. It may also help to form the stalk by which the indusium is connected to this ring, but the development of this was not followed out in detail in the later stages and therefore no definite statement can be made. The periblom of both these sections develops very slightly as far as followed, and as part of section V is pushed around under the base of the sorus (Fig. 53)

apparently as follows in the formation of the soralia on the indusium. The ventral portion of both these sections grows very rapidly in a radial direction, keeping pace with the lengthening of the sorus, and giving rise thus to a part of the indusium surrounding the latter (Figs. 51 - 54), becoming finally more than half the width of the capsule (*i.ind., o.ind.*, Fig. 55). The outer end of these layers later spread out peripherally to meet the ventral end of section VI and thus enclose the cells of the sorus. This portion of each of these two sections becomes several cells in thickness (Figs. 54, 55), and gives rise to a part at least of the ventral portion of the gelatinous ring, while the inner portions (*o.ind., i.ind.*, Fig. 54) remain one cell thick even till maturity. This latter statement, however, is strictly true only of the basicopic portions of these segments, as we shall see later.

The Sporangia.

It remains to describe the development of the most important division of the soral segments, the basicopic ultimate marginal cell. This has a very interesting and significant history, since each is the mother cell of all the sporangia of the sorus formed in its segment. It is the "sorus mother cell" of Bässig, but as it is not the

is only a little longer than to a part of the incusium as well, this name is not strictly appropriate. There is, as we shall see, no single cell after the two tertiary marginal cells are formed which gives rise to the placenta and sporangia only, nor to these and the whole of the indusium. No true dermatogen wall is ever formed in this marginal cell, hence the placenta and sporangia are not of strictly epidermal origin. They are formed rather, as was found in the origin of the sporocarp from the leaf, from a cell that is capable of forming at least two meristem layers, as it actually does in the segments of the stalk (Fig. 50).

In the growth of this part of the capsule that takes place soon after section VI is formed, the marginal cell elongates in a radial direction. At the same time its wall begins to separate at the outer or ventral end from the outer cells of section V (s.c., Fig. 51). Meanwhile the marginal cell has grown in the direction of the length of the sporocarp also (Figs. 52, 64). It soon splits by a transverse anticline into halves, of which the acroscoptic one soon comes to be the larger. Then each of these divides by another transverse anticline (Fig. 65) forming thus four cells, of which the basiconic cell of the first pair formed in the acroscoptic cell of the first pair soon becomes the largest (para-spermoc., Fig. 64), and since

the primary macrosporangium cell or the macrosporangium of the sorus, we shall call it the primary macrosporangium mother cell. The sister cell of this latter divides once more by a transverse anaphysis (Figs. 51, 55) and thus five cells are formed from the basicopyle marginal cell, two on each side of the primary macrosporangium mother cell. Of these the one next to the latter on each side (p.m.s.p.m.c., Figs. 52, 54) is a primary microsporangium mother cell, each giving rise to all the microsporangia of the sorus on its side of the placenta. The outer cell on each side (i.ind., Figs. 52, 54) forms the inner layer of the lateral part of the indusium. These five cells are at first much alike as they appear in a transverse section of the capsule (Fig. 51), but the contents of the three middle ones soon become much denser (Fig. 57).

In horizontal section it is soon seen that while the outer or indusial cells, or the five, remain in contact with the cells of section V, the three middle or sporangium mother cells are separated from this by a narrow slit (s.c., Fig. 57). This is the beginning of the soral canal which we have already noticed in the cross section (s.c., Fig. 51). This cavity constantly increases in size by the pushing out of the indusial cells on each side of the sorus and thus forcing apart some of the cells of section V from the sporangium mother cells.

During the development of the spore case, the basal cell of the marginal cell (the macroscopic marginal cell), usually by a transverse anticline (Figs. 52-53, 55), into two cells, one of which helps to form the outer layer of the indusium in its own segment, while the other goes back to form the sorus of the next younger segment (*o.ind.*, Figs. 56, 58).

Returning now to the development of the five cells formed in the basiscopic marginal cell as seen in a cross section of the capsule, we find that at first it is very similar for all, and we may take for further study, sections in the plane of the macrosporangium mother cell. This is seen to elongate radially and then to divide by a periclinal (Fig. 52), then by further growth and division of both of the cells thus formed (ma-sp.m.c., Fig. 53) there arises a row of seven or eight cells receding from about the center of the capsule nearly to the ventral surface (Fig. 54), all of which are separated by the soral cavity from the cells of the inner layer of the indusium arising in section V. From the increase in size of both of the cells formed from the marginal cell by the first periclinal, from the occurrence of nuclear spinules in both, and from their relation to the surrounding cells, there can be no doubt that both divide further and give rise to

of the microsporangium and the capsule of the sporangia. None of the latter then can be developed in cells in capsular dorsal or inner initials Basgen ('0) though possible. In sagittal section (Fig. 5) it is seen that, as suggested above, periclinal walls are formed in the microsporangiiferous mother cells and the inner initials cells upon opposite sides in the macrosporangiiferous mother cells, as they keep pace with those in initial growth. By the time that the whole number of macrosporangiiferous mother cells has been formed in the scutus, the soral canal has been away from the median wall below and cliptilis toward the median wall at the ventral end (Fig. 5'). It is sometimes the case as Basgen has pointed out that the sporangial cells may grow over into contact with the inner layer of the inusium, but there is certainly no growing together after the first separation and the phenomenon has no significance. Soon after the soral canal has taken this curved form the cells of sections V and VI come into contact and thus close the outer end of the canal. They soon fuse together firmly but the line of junction remains visible for a long time, (fig. 5').

While the five divisions of the basicone are still cells have increased in numbers by radial growth they have also been developing in other directions. The soral canal is first conical but soon becomes very

(s.c., Figs. 1, 2), while the microsporangium mother cells remain in their original positions (Fig. 1). The microsporangium mother cells swell out in a triangular shape (mi-sp.m.c., Figs. 1, 2), so that one microsporangium mother cell is still so large that its隔壁的 size. For this reason sagittal sections will illustrate the median wall better than the one shown in Fig. 27, which does not show anything of the microsporangium mother cells, but simply the soral cavity and the two layers of the lining. On each side of the microsporangium mother cells, the microsporangium mother cells might thus easily be overlooked.

As the microsporangium cell grows and swells like a swelling like form, with a rounded outer end, it is first pushed out into the soral cavity, by the growth of the pluronic cells of the acroscopic part of the basiscopic half of section VI (par.b., Figs. 28, 29). Meanwhile the microsporangium mother cell on each side has increased in thickness and divided by an inclined parallel segment wall (mi-sp.m.c., Fig. 29), when each of the resulting cells divides by a similar wall (Fig. 31) and thus four cells are formed on each side of the microsporangium mother cell. These are pushed apart by the growth of the microsporangium mother cells so as to form narrow right angles of their original line (Fig. 32). The

form a large longitudinal cavity in the apical cell of the microsporangium.

The microsporangium is formed from the apical cell of the plane apical meristem of the apical apical cell division when (in sp., Figs. 51, 51). The first cell formed is seen in a cross section of the capsule is small toward the ventral side of the mother cell (Fig. 51). There is no remains of old or young cells from the former in the sorus as described by Bissgen ('10) and as figured in Pilularia by Müller ('37), but these lie squarely against each other (Figs. 52, 51). The apical cell of the microsporangium cuts off two small rhombic sides which form the lateral and basal walls of the sporangium and that part of the placenta or stalk which lies between the sporangium and the region of the placental bundle (Fig. 53). Then a pariclin is formed near the outer end of the apical cell and the archesporium is therefore (are, Fig. 52) similar to the sporangiophore wall developed, the former giving rise to Russow's "shells" of the microspores.

While the microsporangium has been developed, there is a marked change in the cells derived from the microsporangium. A cell in the apical section VI (Fig. 51) is visible in between Figs. 51, 52 and 53. It is a large cell, the nucleus being large. It is

inches long, and divided into two rows of small rounded cells at first. These are arranged in four rows, or approximately in three, two cells apart, on either side in the length of a sorus of eight microsporangia. There would thus be four cells derived from a microsporangium in a longitudinal section of a capsule 17½ older than that shown in Fig. 26. Each of the cells arising from that one of the original four nearest the microsporangium (mi-sp., Figs. 21, 22) swells away from the placenta and divides into a basal cell, and an outer cell which gives rise by inclined walls to the tetrahedral apical cell of a microsporangium; while the cells derived from the original four furthest from the microsporangium, form the outer cells of the placenta in that region (pa., Figs. 21, 22). The basal cell of the microsporangium can perhaps be considered as homologous with the stalk cell formed in the sporangia of many of the heterosporous Lycopodiophytes, but there was nothing seen in the development of the microsporangia that could be regarded as such, and Marsilia thus differs from Pilularia, in which Campbell ('3) has described such a cell as being characteristic.

The cells derived from the lower class of section VI as described above, consisting of four rows, giving rise to two rows of cells, is seen in horizontal section (Figs.

0 - ..), while the outer ones are placed in the inner cells on the side away from the microsporangium, the cells. From certain of these basal cells, in a manner described in detail later, are formed the placentule bundle and the placental branch.

Meanwhile the inner and outer indusial cells, on both sides of the sori, have kept pace with the growth of the sporangial cells. The acroscopic part of section V and the acroscopic marginal cell have each split by a transverse anticline (Fig. 75) to form the lateral parts of the outer layer of the imusia of the adjacent sori, and these, in connection with the cells of section II, complete the outer indusial layer, surrounding the sori through its whole length (o.ind., Figs. 71, 70 - 72). The lateral inner indusial cells, arising from the basocentric marginal cell, together with the cells of the basocentric half of section V complete the inner layer of the indusium also. Each of these layers contains only a single layer in thickness even at maturity, and division walls are formed in their cells except those terminating on the sori, and by growth in this manner the thickness constantly increases. In spite of the sorus evagination, the sori are pushing out into air (Figs. 70 - 72). During the growth of the indusial cells intercellular spaces appear, between the two layers, and they increase

from the primary cell, and the latter from the secondary cell. The primary cell, which originates from the mesoderm, is the larger of the two, and contains the nucleus (see Fig. 1). Following the division of the primary cell into two smaller cells, the nucleus continues to divide, and the secondary cell gives rise to a number of small daughter cells (Fig. 2). By this time also the secondary cells begin to encapsulate the primary cell in a thin capsule of opposite side (Fig. 3). They then appear to be situated on opposite sides of the capsule, and at this stage they have (Fig. 4) they are also alternate in size, one being large and the other small, and originate from different parts of the primary cell. As a result of the continued division on both sides of the capsule.

The primary microvilli in the primary cell are shown on the sides of the primary nucleoplasm in the mother cell, and are similar to those described by Ringer in his first article on microvilli and motor cells. Hence they can not arise from strands of the apical cell as suggested by Dr. Ringer and Brugge in their article. The placental blood has also a similar condition, but from the time when the secondary cell has apical cell to the time when Ringer and Ross describe, not very long afterwards, it arises from the basal cell, and was formed from the microvilli seen in section VI.

We now turn to the posterior commissure, which is the second of the two main centers of the trigeminal nerve, and described in connection with the development of the root, in order to avoid unnecessary complication. The ventral portion of each lateral bundle arises from the lower half of the pons in the median sulcus between the inner margin of the corpus and the dorsal bundle, situated between its two branches as has been shown by Russow. Two of these combine in the course of the single bundle near the point of the capsule (l.b.r., Fig. 57, 58), while the other remains separate, turning toward the median joins the ipsilateral bundle (p.b.r., Figs. 57, 58, 59). Of the two branches of the lateral bundle, one arises in the mesencephalic or the aeroscopic half of the same segment (l.b.r., Figs. 57, 58), while the other arises from the aeroscopic quarter of the aeroscopic half of the next older segment (l.b.r., Figs. 57, 58, 59). These two branches form with the marginal cell a small individual, but the ganglion cell forming the cellis of Russow (Fig. 59), so known in terminology. In some (l.b.r., Figs. 57, 58, 59) these three are represented by a single, very large

and Fig. 72 (L. M., Fig. 70-71), we notice another division of the capsule. There are two groups of cells situated on either side of the central layer of the stroma. These are less regular than the columnar vascular chiasmata. The cells lying between them (Fig. 72).

It has been shown above that the portion of the histoscopic half of section VII divides into two portions (Fig. 77) and that the acrosopic one of these contains the median cells of the placenta (p. 4, Fig. 72). Most of these cells form a part of the general parenchyma of this region of the placenta, but several rows of these cells at the point between the opposite microsporangia are in contact with the first segment of the apical cell of the microsporangium, give rise to the placental bundle, which runs through the whole length of the placenta. In a region about opposite the inner ones of the sorts (p. 4, Fig. 71, 71') these subcells also become specialized to form the placental branch which runs across the whole breadth of section VI from the single origin of the lateral bands to the placental bundle (p. 4, Figs. 71, 71').

The sterile Setae of the Capsule.

We come now to the consideration of the capsule.

at the base of the capsule lining, and the small segments, to its position on the dorsal midline. In section I the ventral nerve is also found dorsal to the midline, also dorsal and lateral branch, located in the pleurae of sections III and IV as in the small segments (Figs. 11, 12). No trace of the placodal strand or precentral bundle is seen here, and in the other two sections no lateral pair is developed. The pleromes of sections II, V and VI, in the young's segment of the thorax, and probably that of all the sections except section I of the middle segment, is apparently devoted to the formation of the basal portion of the goldenous ring (i.e., Figs. 13, 14). In the oldest or basal segment of the thorax the plerome and periblast of all the sections are confluentified to form the basal half of two layers of thickened cells (b.w., Figs. 15, 16), lining the hypodermal cells of the capsule wall, the stretching across from the dorsal to ventral midline. In the peripheral of section I in the youngest segments, the plerome is the first to appear, and in this also, there is developed a pair of "niches" or "lens-shaped cells" of Ewer (i.e., Fig. 17). Into a small opening at one margin of the latter object a cavity containing the cells

S = $\frac{1}{2} \pi r^2$.

the *Leptostoma* come from the epidermal cells, some of which, as we have seen, divide obliquely, and the basal and uppermost ones divide longitudinally, the two sections thus giving rise to a basal cell of the segment. Each of these divisions forms 1/4 one, periblennial, and between layers, in the basal and upper parts, it commences from the plerome of the first four segments. At each pinnule there is a periblennial duct, which, in a hair-follicle, and the partitions between those, which are derived from the plerome, have many intercellular openings through them at maturity. Trichomes are formed from the epidermal cells in a very regular arrangement. Certain rows of the periblennial cells become specialized as trichome reservoirs. The pinnules or divisions of the leaf arise as a result of the continued activity of the marginal cells of certain segments, and their limits do not correspond exactly with those of the segments.

The sporocarp arises from a single ultimate cell in 1 cell of the second segment of the leaf, as a lateral branch of the latter, which consists of only six or seven segments on each side. The second sporocarp is all active in the same way from one of the basal marginal cells of the first one, and is like a secondary branch of the leaf. The apical cell is however, in position, in the middle of this compound branch, and it bears a very large

The semicircular zones of the capsule are formed by longitudinal anastomoses in a six-sided arrangement around the marginal cell of the sphaerule zone. The basal portion of the capsule is shown enlarged in the figure of section I, the lateral bundles in both sections III and IV and the placental bundle and border in section VI. The epidermis is of one layer of cells and the hypodermis of two, both layers of the latter consisting of very thick-walled cells. The trichomes of the epidermis and the canals within the hypodermis are like those of the actiole, except that the latter are smaller and more irregular. The sporangia of each sphaerule are all derived from a basiscopic ultimate marginal cell, which gives rise also to the cells forming the inner layer of the lateral portion of the indusium. The single primary microsporangium mother cell and the two primary megasporangium mother cells of each sphaerule are sister cells, formed by the first division of the marginal cell. They are thus superficial, but not strictly epidermal, in origin. The sphaerule canal arises by a splitting between the two primary sporangium mother cells and follows the cells of section V, beginning at the ventral margin of the capsule and extending laterally to the inner edge of the dome. From the dome section it extends

the leaf cell, as a result of which the primary section cell, which forms also the primary segment of the leaf cell, A stalk cell is formed in the development of the microsporangium, but nothing analogous with this was seen in the case of the macrosporangium. The indusium of such sporangia arises partly within its own segment, but part also from the next older segment. It is derived from the dermalogen cells of sections II and V and from the marginal cells. The gelatinizing tissue of the dorsal part of the capsule arises largely, if not entirely, from the plerome of section II; that of the ventralside from the dermalogen of sections II and V, and that at either end of the capsule probably comes from all three meristematic layers.

Conclusions.

The leaf of *Marsilia* agrees closely with those of most of the Lepidosporangiata. Ferns which have been compared, in its origin, in its growth on a two-sided leaf cell, and in the development of the lamina by the peripheric activity of the marginal cells of certain of its soriusses. The position of the primary or section cells in the soriusses of the leaf, will remain clear when

but found in Asplenium *Scandens* Spreng. (1825), is very different from the description given by C. L. P. (1821) for the *Leptosporanum* in general, so far as one can judge from the description alone. It is very evident that, as Bower has suggested, there is great need of additional work on the origin of the distinct regions in the various organs of the Ferns. Many of the accounts at present available, of the development of the rachis and laminae are very unsatisfactory because of the lack of details in both descriptions and figures.

The sporocarp is certainly to be considered as a branch of the leaf, since it is derived from the apical cell of the leaf. The second sporocarp is usually a branch of the first and hence a secondary branch of the leaf, while the third may in like manner be a tertiary branch. In the shape of its apical cell and in the way it is cut off from this, the sporocarp agrees exactly with the leaf, while in the primary divisions of the sporocarp, it differs from the leaf only in the intercalation or in exertion, see section dorsal to the marginal cell. The capsule, so far as is at present known, is probably a continuation of the leaf itself. There is no indication from this source that the sporocarp contains anything homologous with the lamina of the leaf. The outer protective layer of the sporocarp is probably a continuation,

the capsule to the bone, and the bone to the muscle. We may, however, consider the capsule as being composed of two parts, the fibrous layer, which is well seen in position of connection.

It is difficult now to assess the importance of G. 1. The capsule of M. adductor pollicis is said to be placed together with the muscle tendon at the ventral margin of the capsule, is to be anatomized. The cause of development is given above. The same difficulty is of course found in the interpretation of Russow and Bⁿ. In the figure the capsule is said to be loose, and in that of Mauduit and Campbell the capsule is a pinnate band with the radialis at the dorsal margin and with a pinnate loose sortie.

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Explanation of Figures.

Abbreviations used :- A. direction of the rays; a.c. axial bundle of trichomes; ac. aciculus; an. anestomosis; as.w. acrocapitile sphaerule; B. direction of base; b.c.b. basal coil of trichome; b.m.c. basal marginal cell; Br. branch, (axial bran.); c.a. column of all cells; c.s. central sphaerule; c.s.a. a circosporangium cell; b.c. basal coil of capsule; C. cells - c.p. transverse partition; c.p.c. transverse partition cell; c.p.o. pores in transverse partition; D. dorsal view; d. division; I. II. loc. macrocysts in sections I, II; d.c. dorsal canal or sporocyst; d.v. dorsal view canal; d.v.s. dorsal view of sporocyst; E. sporocyst; F¹. first sporocyst; F². second sporocyst; F_{m.c.} macro cells of sporocyst; f.t. laminous film; h.n. halvinian folia in section I; h.v. halvinian; ind. in older; i.ind. inner older; in- older; i.s. intercellular space; i.s.e. intercellular space; L. lamellae; L.c. longitudinal section of spores in section V; l.m. lateral margin of section V; l.m.l. longitudinal margin; l.m.s. longitudinal section; l.m.v. longitudinal view; l.s.c. longitudinal section of spores in section V; l.v. longitudinal view; l.v.s. longitudinal section of spores in section V.

Plate I.

Fig. 1. A nearly vertical section of the stem apex showing the surface of the stem and the leaf mother cell with one segment wall. x300.

Fig. 2. Transverse section of stem from apical cell of young leaf. x300.

Fig. 3. Section of stem between vertical and horizontal planes, showing surface of stem apex and transverse section of leaf. x300.

Fig. 4. Sagittal section of a leaf nearly at the end of apical growth. x200.

Fig. 5. Part of a sagittal section of the petiole of an older leaf. x100.

Fig. 6. Ventral surface of tip of an older leaf. x100.

Fig. 7. Dorsal surface of the tip of a similar leaf x100.

Fig. 8. Surface of the tip of a very young leaf showing in the edges of the two young segments on one side and the apical cell surface some chlorophyll and air bubbles still unbroken. x300.

Fig. 9. Hull of a nearly transverse section of a young leaf, showing the single cell membrane separation of the disc section. x100.

Fig. 10. A similar section showing the same membrane. x100.

Fig. 11. A transverse section of a still older segment. x10.

Fig. 12. The same still older. x10.

Fig. 13. Transverse section of a still older segment. x10.

Fig. 14. Transverse section of a petiole showing an intimate arrangement of the first grade. x750.

Fig. 15. Similar section showing the intimate arrangement of the sixth grade. x1750.

Fig. 16. Transverse section of petiole in which the dermal and hypodermal layers are confluent and the periderm cells are nearly ready to cut off the cortex. x750.

Fig. 17. Part of similar section of a still older petiole. x100.

Fig. 18. The same section of a still older petiole. x100.

Fig. 19. Transverse section of a nearly mature petiole. x10.

Plate II.

Fig. 20. Part of a transverse section of a petiole. x100.

Fig. 21. Tangential section of petiole showing the early periderm. x100.

Fig. 24. The same as Fig. 23, $\times 60$.

Fig. 25. The same as Fig. 23, $\times 100$.

Fig. 26. The same as Fig. 23, showing the beginning of the longitudinal partition. $\times 100$.

Fig. 25. Surface view of the longitudinal partition showing the fibers in a normal arrangement. $\times 100$.

Fig. 27. Sagittal section of particle, showing a portion of a longitudinal fiber of Fig. 1. $\times 500$.

Fig. 27. Surface view of particle in the same position showing the arrangement of the trichomes. $\times 10$.

Fig. 28. Leaf of *Lycopodium*. $\times 10$.

Fig. 29. Normal surface view of leaf. $\times 100$.

Fig. 30. Near the midrib. $\times 10$.

Fig. 31. Horizontal section of leaf, showing the beginning of the fibrous tissue. $\times 100$.

Fig. 32. The same leaf, showing origin of terminal pinnules. $\times 100$.

Fig. 33. Longitudinal section of a leaf, showing the well developed pinnule. $\times 100$.

Fig. 34. Transverse section of long fibrous pinnule, of the leaf of Fig. 31. $\times 100$.

Fig. 35. A similar section of a long fibrous pinnule.

Fig. 36. A similar section of a long fibrous pinnule.

Fig. 37. A similar section of a long fibrous pinnule.

Leptostoma sp. $\times 0.$

Fig. 9. A portion of a young sporocarp showing the apical region. $\times 0.$

Fig. 10. The same at a later stage. $\times 0.$

Fig. 10. Median longitudinal section of the young leaves, an older leaf, portion of the sporocarp, fragments, showing the inner surface of a chloride leaf, and cross section of the first sporocarp. $\times 0.$
Isolation of specimens occurs from the first. $\times 00.$

Plate III.

Fig. 11. Transverse section of stem, and a young leaf in two stages occupying all round parallel to the side. $\times 0.$

Fig. 12. Inner side of young leaf with a sporocarp in which the segmentation of the apical cell is nearly finished. $\times 00.$

Fig. 13. An older sporocarp on a pedicel with capsule bent against the wall. $\times 00.$

Fig. 14. Transverse section of a young sporocarp. $\times 0.$

Fig. 15. The same in an older stage. $\times 0.$

Fig. 16. Transverse section of a young sporocarp of the second leaf to be formed. $\times 00.$

Fig. 17. The same in an older stage. $\times 00.$

Fig. 7. The same transverse section of shell, showing the condition of IV in the successive layers of growth, compared with Fig. 7. x 0.

Fig. 8. The same transverse section of shell, showing the condition of V in the successive layers of growth, compared with Fig. 7. x 0.

Fig. 9. Transverse section of shell of the specimen shown in Fig. 1. x 70.

Fig. 11. The same section of specimen, showing the condition of some early, twin, relatively thickened older myonemes. x 70.

Fig. 12. Part of transverse section of shell of specimen x 70.

Fig. 13. The same still older. x 100.

Fig. 14. The same at the time of elevation of shell. x 100.

Fig. 15. The same still older. x 100.

Fig. 16. Ventral surface of cancellous layer of shell. x 70.

Plate IV.

Fig. 17. Posterior median section of shell, showing the condition of IV in the second instance. x 0.

Fig. 18. The same section of shell. x 0.

Fig. 1. Vertical projection of the boundary of the domain of the initial value problem (1.1) for $\alpha = 0$.

Fig. 2. Horizontal section of the boundary of the domain of Fig. 1, near the vertical slice $x = 0$.

Fig. 3. The same, near the boundary of Ω_0 , $x \approx 0$.

Fig. 4. The same, near the left boundary of Ω_0 , $x \approx -0.75$.

Fig. 5. The same, near the right boundary of Ω_0 , $x \approx 0.75$.

Fig. 6. Spherical section through the center of the ball of constant boundary value $\phi = 0$ (Fig. 1, $x = 0$).

Fig. 7. Spherical section passing through III or IV of capsule, near the boundary of Ω_0 , $x \approx 0$.

Fig. 8. The same, near the boundary of Ω_0 , $x \approx 0$.

Fig. 9. Spherical section through the center of Ω_0 , $x \approx 0$.

Fig. 10. Vertical projection of the boundary of the domain of the initial value problem (1.1) for $\alpha = 1$ (near the capsule), $x = 0$.

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V i t a .

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The writer of this biography was born in Cromwell, Conn., on the 1st of April, 1870. He received his early education in the public schools of Cromwell and Middletown Conn., and took his college course at Wesleyan University, in the latter town, receiving the degree of B. S. in 1893. During the college year of 1893-4, he was a graduate student at Wesleyan. Since the fall of 1894, he has been a graduate student at Johns Hopkins University, with Botany as major subject and Zoology, Physiology, and Minors.

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